Genus Vol. 10 (3): 497-522 Wrocław, 31 X 1999

Contribution to the knowledge of Ecuadorian *Pronophilini*. Part III. Three new species and five new subspecies of *Lymanopoda* (\*\**Lepidoptera*: *Nymphalidae*: *Satyrinae*)

Tomasz W. Pyrcz <sup>1</sup>, Keith R. Willmott<sup>2</sup> & Jason P. W. Hall<sup>2</sup>

- Zoological Museum, Institute of Zoology, Jagiellonian University, Ingardena 6, 30-060
  Kraków, Poland, E-mail: pyrcz@zuk.iz.uj.edu.pl
- 2. Department of Entomology and Nematology, University of Florida, Gainesville, Florida 32611, U.S.A., E-mail: krwi@gnv.ifas.ufl.edu

ABSTRACT. Three new species and five subspecies of Lymanopoda are described from Ecuador and their affinities within the genus are discussed. Misidentifications in Brown's (1943) survey of the genus Lymanopoda in Ecuador are corrected. Lymanopoda hannemanni L. Miller, 1991, is synonymised with Lymanopoda confusa F. M. Brown, 1943 (n. syn.), and the female of this species is described and figured for the first time.

Key words: entomology, taxonomy, Lepidoptera, Nymphalidae, Lymanopoda, new taxa, Colombia, Ecuador, Peru, Podocarpus National Park.

# INTRODUCTION

The genus Lymanopoda Westwood, 1851 is a member of the tribe Pronophilini sensu Miller (1968), which was downranked to subtribe Pronophiliti by Harvey (1991), an entirely Neotropical section of the nymphalid subfamily Satyrinae. It comprises approximately sixty species almost exclusively confined to the Andes, with only two representatives in the Central American mountains (Pyrcz, in prep.).

Typologically, the adults of *Lymanopoda* can be characterised by their small to medium size (forewing length 20-25 mm) compared to other members of the tribe, triangular forewings which usually have an acute apex and often a convex

outer margin, oblong hindwings, often with scalloped outer margins and a short tail-like emargination at vein Cu<sub>1</sub>, rather short antennae, approximately 2/5 length of the costa, and eyes which are covered with short, sparse setae. The venation pattern is typical of the tribe *Pronophilini*, with the base of the cubitus of the forewing moderately swollen and the anal weakly so, and the disco-cellular vein of the hindwing between veins M<sub>1</sub> and M<sub>2</sub> sharply angled basally near M<sub>1</sub> (Brown 1943; Miller 1968).

posterior to the ostium bursae. gland (of unknown function, most probably producing an egg gluing secretion) the distal part of the posterior apophysis of the papillae anales and an accessory phied). In the female genitalia, synapomorphies include: a sclerotised lamella on sculptured processes on the valvae (in a few cases the dorsal process is atroof the uncus); a (usually) strongly sclerotised sub-scaphium; and two prominent not homologous with the subunci, appearing to be a modification of the basal part ("pierellization"-type distortion of the ground plan (sensu Schwanwitsch 1925)). Other generic synapomorphies are evident in the male genitalia, including: the (although a weakly sclerotised projection occurs in all species which is probably tegumen at the dorsal junction with the uncus; the complete absence of subunci presence of a superuncus (sensu Razowski 1996), a bulbous projection of the band is broken and displaced in the discal cell, connected to the postbasal band lies beneath, or posterior to, that cell) and the hindwing ventral surface median the remainder (we refer throughout the text to wing cell spaces by the vein which synapomorphies of the genus Lymanopoda can be identified in the wing pattern: the ocelli in forewing cells Cu, and Cu, are always displaced basally in relation to from white, dull brown to russet, metallic silver, green and blue, at least two Despite the coloration of the wings varying greatly between species, ranging

As far as is known, the larvae of Lymanopoda feed on Chusquea bamboo in cloud forest (Schultze 1929; Adams 1985), or Swallenchloa bamboo (Poaceae) in the páramo (Pyrcz unpubl.), and only exceptionally on other gramines (L. caeruleata Godman & Salvin, 1880, a species endemic to the Sierra Nevada de Santa Marta, Colombia, was observed by the senior author while laying eggs on Bambusa). The larvae of only one species, L. samius Westwood, have been described (Schultze 1929), and therefore no comparative taxonomic characters are available as yet from the immature stages.

The species of Lymanopoda show intricate patterns of horizontal and vertical distribution. Whereas one species, L. obsoleta (Westwod, [1851]), is nearly Panandean, most have a much more restricted distribution, several being single range endemics, including L. confusa F. M. Brown 1943, discussed in this paper. They are found in premontane forests from around 800 m (L. panacea (Hewitson 1869)), in cloud forests and up to boggy páramo over 4000 m (L. huilana Weymer 1890), within well defined, sometimes very narrow bands of elevation (Adams 1985; Pyrcz & Wojtusiak in press). While L. obsoleta occurs from approximately 1800 to 2900 m (Adams 1986; Pyrcz & Wojtusiak in press), L. marianna Staudinger, 1897 is known from Venezuela from a narrow band between 3000 m

and 3200 m (Adams & Bernard 1981). The patterns of altitudinal distribution appear to be related to ecological specialisation and interspecific interactions (Pyrcz & Wojtusiak in press). The species with particularly narrow vertical ranges are in most cases exclusive inhabitants of the cloud forest - páramo ecotone, such as *L. marianna* and three of the species described herein. The adults of cloud forest *Lymanopoda* are strongly attracted to decomposing organic material, including carrion (Willmott & Hall unpubl.), dung, urine, fruits, and mineral matter found in mud (Adams 1985, 1986), but páramo species have not yet been reported to be attracted to baits. Páramo species are energetic butterflies, flying low above the ground, zigzagging among *Espeletia* composites and active only during longer periods of sunshine, whereas the cloud forest species are less motile, usually not moving far away from stands of their *Chusquea* hosts; individuals can be observed for several consecutive days in the same spot.

Brown (1943) surveyed the genus Lymanopoda in Ecuador, describing two new species and listing a total of 11 species for the country. That paper contains a number of identification errors resulting from the fact that Brown had no access to English and German type material, and is now largely outdated due to more extensive sampling for montane butterflies during the past decade in some of the more remote areas of the country by several lepidopterists. Nevertheless, it provides a point of reference from which to begin faunistic, revisional and taxonomic research. The first author is currently working on a revision of the entire genus Lymanopoda, while Keith Willmott and Jason Hall have been working since 1993 on the taxonomy, ecology and biogeography of the entire true butterfly fauna (Papilionoidea) of Ecuador. Therefore in this paper we describe the new taxa within the genus Lymanopoda that have come to light during our studies of Ecuadorian pronophilines. The following acronyms are used throughout the text:

AMNH: American Museum of Natural History, New York, U.S.A.

BMNH: The Natural History Museum, London, United Kingdom;

MALUZ: Museo de Artrópodos de la Universidad del Zulia, Maracaibo, Venezuela;

MNCN: Museo Nacional de Ciencias Naturales, Quito, Ecuador;

MUSM: Museo de Historia Natural de la Universidad Nacional Mayor de San Marcos, Lima, Peru;

MZUJ: Muzeum Zoologiczne Uniwersytetu Jagiellońskiego, Kraków, Poland;

PUCE: Pontificia Universidad Católica, Quito, Ecuador; ZMHU: Zoologisches Museum, Humboldt Universität, Berlin, Germany;

AJ: Collection of Artur Jasniski, Warsaw, Poland;

KWJH: Collection of Keith WILLMOTT and Jason HALL, Gainesville, Florida, U.S.A.;

MB: Collection of Maurizio Bollino, Milan, Italy;

PB: Collection of Pierre BOYER, Le Puy Sainte Réparade, France

TWP: Collection of Tomasz Pyrcz, Warsaw, Poland

# LYMANOPODA FROM ECUADOR

# Lymanopoda ichu Pyrcz, Willmott & Hall n. sp. (Figs 1 & 20)

### DIAGNOSI

shape, proportions and sclerotization of some main structures, especially the which is V-shaped when viewed posteriorly, which is noticeably more deeply dark brown, not orange-brown. The genitalia (Fig. 20) are devoid of a dorsal of being incurved as in L. excisa browni, and by the basal area of the ventral dian black dots which form a row parallel to the outer margin in L. ichu, instead straighter outer margin below the apex, by the hindwing ventral surface postmeexcisa browni n. ssp. (described below) by the less acute forewing apex, the valvae. The individual illustrated (Fig. 28) corresponds to a population found on are ended distally by a series of spines. L. melia is exceptionally variable in the L. huilana and L. melia differ from both L. ichu and L. caracara in that the valvae cleft in L. caracara. The valvae are also slightly more elongate in L. caracara 28). Both L. ichu and L. caracara share a hooked uncus, the dorsal surface of L. huilana huilana Weymer, 1890, (Fig. 27) and L. melia Weymer, 1911, (Fig. process on the valve and in this respect are similar to L. caracara (Fig. 18), forewing apex, paler dorsal ground colour, and has the base of the forewing costa L. caracara n. sp. (described below) is also similar but is larger, has a more acute forewing being black except for a small brown marking in the discal cell. the western slopes of the Central Cordillera in Huila (Colombia). This species is distinguished from the superficially similar Lymanopoda

# DESCRIPTION

surface pale cream. Wings: forewing (length: 18-20 mm, mean: 19.2 mm, n=3) surface of club dark brown. Thorax: dorsal surface dark brown, ventral surface ventrally pale brown with few dark brown hairs; antennae 1/2 length of costa, with sparse setae; labial palpi dorsally dark brown with few pale brown hairs relation to others. Hindwing dorsal surface dark brown with tiny, faint postmemedial series of white dots, those in spaces Cu, and Cu, displaced basally in elongate, distal margin smoothly rounded; hindwing elongate and rounded. pale brown; legs pale brown. Abdomen: dorsal surface dark brown, ventral brown and very sparsely scaled with white only at base of each segment, dorsal along outer margin; five white dots reflected from dorsal surface, those in Cu<sub>1</sub>-M<sub>3</sub> dusted with ochreous along costa, on apex distally as white subapical dots and dian white dots in spaces Cu<sub>2</sub>-M<sub>3</sub>. Forewing ventral surface ground colour black, Forewing dorsal surface dark brown; basal third of costa orange-brown; a postmargin, except in lower half of discal cell; a series of black postmedian dots scales forming a poorly defined darker band stretching from base towards outer ringed with black. Hindwing ventral surface ochreous, suffused with chestnut parallel to outer margin, in spaces 1A+2A-M<sub>1</sub>. Male genitalia (Fig. 20): valvae Male (Fig. 1): Head: frons with tuft of dark brown hair; eyes brown, covered

lacking upper process; superuncus well developed; uncus hooked with shallow dorsal groove; aedeagus smooth.

Female: unknown.

#### LIFES

Holotype male: Ecuador: Loja: km. 20 Jimbura-San Andrés rd., 3300 m, 24.IX.1997, K. R. WILLMOTT leg., to be deposited in BMNH. Paratypes: 2 males: same data as holotype, in KWJH.

# ETYMOLOGY

This species is named after the Quechua word "ichu", which is used to designate the bunch-grass typical of puna and páramo grassland in southern Ecuador and Peru --the habitats of this species.

#### REMARK

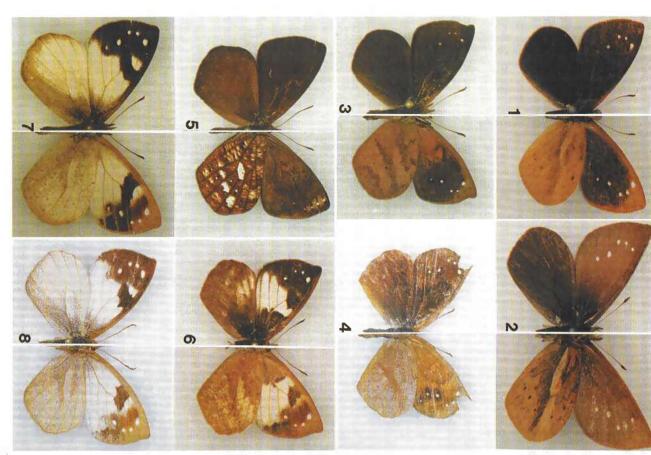
L. ichu n. sp. is currently known only from the type locality in Ecuador. It is closely allied in facies to an undescribed species (Pyrcz, in prep.) occurring in the area of Chachapoyas in northern Peru, with which it shares similar wing shape and ventral surface colour pattern but differs considerably in the male genitalia. As for other Ecuadorian species, there are strong affinities with L. caracara n. sp., found further north, as indicated by genitalic morphology, the elongate wing shape, the shape of the row of postmedian black dots of the hindwing ventral surface and the similar habitat type and altitude L. ichu is also related to L. melia on the same morphological and ecological grounds, even though L. melia is distinctly marked, being predominantly white. L. ichu, L. caracara and L. melia belong to a group including also L. huilana and L. tolima Weymer, 1911, whose diagnostic feature is that the postmedian black dots on the hindwing ventral surface form a row parallel to the outer margin.

L. ichu was found only at a single site along the Jimbura-San Andrés road, at a steep, small landslide through a pocket of elfin cloud forest surrounded by páramo. This landslide had extensive bamboo secondary growth, and individuals were found flying just above the surface of the bamboo during a long period of bright sun.

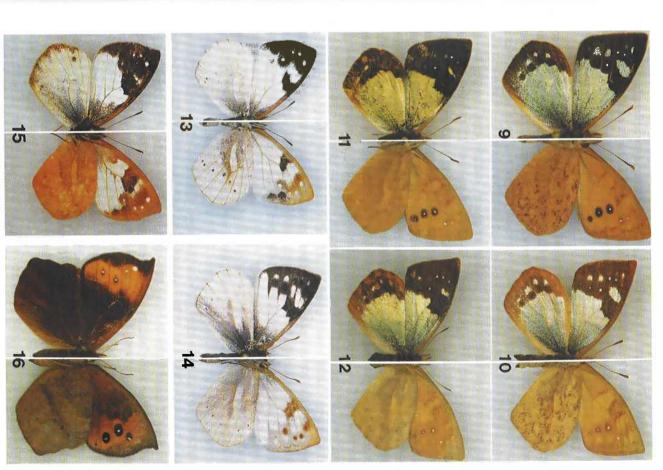
# Lymanopoda caracara Pyrcz, Willmott & Hall n. sp (Figs 2 & 18)

### )IAGNOSIS

This species has the same elongated fore and hindwings and overall hindwing ventral surface colour pattern as *L. huilana* and *L. tolima*, but is immediately distinguished from these species by being entirely chestnut on the dorsal surface. It is also similar to *L. ichu* n. sp., described above (see Diagnosis under that



1. Lymanopoda ichu n. sp. male, dorsum/venter; 2. L. caracara n. sp. male, dorsum/venter; 3. L. excisa browni n. ssp. male, dorsum/venter; 4. Lymanopoda excisa browni n. ssp. female, dorsum/venter; venter; 5. L. nadia n. sp. male, dorsum/venter; 6. L. labineta piniasi n. ssp. male, dorsum/venter; 7. L. nivea bingo n. ssp. male, dorsum/dorsum; 8. L. nivea bingo n. ssp. female, dorsum/dorsum



9. Lymanopoda hazelana summa n. ssp. male, dorsum/venter; 10. L. hazelana summa n. ssp. female, dorsum/venter; 11. L. hazelana hazelana male, dorsum/venter; 12. L. hazelana hazelana female, dorsum/venter; 13. L. nivea nivea male, dorsum/venter; 14. L. melia male, dorsum/venter; 15. L. nivea bonita n. ssp. male, dorsum/venter; 16. L. confusa female, dorsum/venter;

species). It differs genitalically from L. ichu by having a more deeply cleft uncus and a more elongate valva, and from L. huilana and L. melia as specified in the diagnosis of L. ichu.

# DESCRIPTION

suffused with darker brown in basal half and along vein M3, except in discal cel surface of club dark brown. Thorax: dorsal surface dark brown, ventral surface ventrally pale brown with few dark brown hairs; antennae 1/2 length of costa, distal end of discal cell. Hindwing ventral surface ground colour ochreous, outer margin; five white dots reflected from dorsal surface; ochreous marking as ous scales along costa, apex distal of white subapical dots and narrowly along medium brown. Forewing ventral surface dull brown, slightly dusted with ochrehalf, a postmedial series of white dots in spaces Cu2-M1, those in spaces Cu2 and and rounded. Forewing dorsal surface medium brown, slightly lighter in distal elongate, distal margin slightly rounded, apex pointed; hindwing very elongate surface pale cream. Wings: (length: 22-23.5 mm, mean: 22.75 mm, n=2) very pale brown; legs pale brown. Abdomen: dorsal surface dark brown, ventral brown and very sparsely scaled with white only at base of each segment, dorsal with sparse setae; labial palpi dorsally dark brown with a few pale brown hairs, deep dorsal groove; aedeagus smooth. ing upper process; superuncus well developed; uncus strongly hooked and with postmedian dots parallel to outer margin. Male genitalia (Fig. 18): valvae lackwhich is pale yellow in lower half and orange-brown in upper half; a row of black Cu, displaced basally in relation to others. Hindwing dorsal surface uniform Male (Fig. 2): Head: frons with tuft of dark brown hair; eyes brown, covered

Female: unknown.

#### Types

Holotype male: Ecuador: Morona-Santiago, Gualaceo-Chiguinda rd., east of pass, 3300 m, 20.XI.97, K. R. Willmott leg., to be deposited in BMNH. Paratypes (6 males): 1 male: same locality data as holotype, I. Aldas Villafuerte leg., in KWJH; 3 males: same locality and collector, in TWP; 2 males: Azuay: Gualaquiza-Cuenca rd., Sigsig, 1998, 3000-3600 m, P. Boyer leg., in PB.

# ETYMOLOGY

This species is named with reference to its distinctive, strongly hooked uncus, reminiscent of the beak of the Carunculated Caracara, a predatory bird inhabiting the windswept Andean highlands where this species flies.

#### REMARKS

L. caracara n. sp. is most closely related to L. ichu, L. huilana, occurring in northern Ecuador and south-central Colombia, and L. tolima (considered a subspecies of L. huilana by Adams (1986)), endemic to the Tolima massif in the north-central part of the Central Cordillera in Colombia. L. caracara was found

at a single point where the Gualaceo-Chiguinda road traversed a very steep hill covered with low highland vegetation just below paramo, with a noticeable absence of bamboo, where males were encountered flying rapidly up the slope in bright sun.

# Lymanopoda nadia Pyrcz, n. sp. (Figs 5 & 17)

## DIAGNOSIS

L. nadia n. sp. differs from the most closely related species, L. labda Hewitson, 1861, in having four subapical white dots on the forewing ventral surface, instead of three as in L. labda (and also L. lebbaea C. & R. Felder, 1867). It is also darker on the dorsal surface and ventral surface, making the ventral silvery pattern more contrasting. The two species differ genitalically, the valvae in L. labda being more elongate and the dorsal projection smaller in relation to the ventral projection. Genitalic differences are consistent throughout the ranges of both species. L. rana Weymer, 1911 (Peru) is also similar to L. nadia, but has a wide rufous area on the forewing ventral surface and very distinct genitalia.

### ESCRIPTION

subapical area, base of discal cell and most of area between costa and discal cell; ground colour red-brown with paler silvery brown streaks in basal half of space subapical dots in spaces  $R_s$ - $M_s$  and two submarginal white dots in spaces  $Cu_2$  and pale grey. Wings: forewing (length 19-22 mm, mean=20.16 mm, n=29) trianguhair; tibia grey, femur chestnut. Abdomen: dorsally blackish brown, ventrally segment, club flattened costally, blackish brown except last segments chestnut. costa, chestnut dorsally and ventrally, with sparse white scales at base of each ventrally and black hair dorsally, black on third segment; antennae 2/5 length of short, sparse setae; labial palpi twice length of head, covered with grey hair with two short, broad processes of approximately equal size, each with severa basal to this band in remainder of wing. Male genitalia (Fig. 17): valvae broad spaces, distal to postmedial band of silver markings in spaces 1A+2A and Cu2, pupils in spaces 1A+2A-M2, double in 1A+2A and single in remaining cell postmedial band from space 1A+2A-M3; six postmedial black spots with white irregular silver markings filling distal quarter of discal cell and forming a broken 1A+2A, basal area of space Cu2, submarginal area of spaces 1A+2A-Cu1, in Cu, displaced basally in relation to subapical dots. Hindwing ventral surface brown, red-brown scales in discal cell and along distal margin at apex; four white hindwing dorsal surface uniform dark brown. Forewing ventral surface dark Thorax: dorsally blackish brown, ventrally pale grey, covered with short, sparse long "teeth"; aedeagus smooth lar, outer margin slightly incurved below apex; hindwing angular. Forewing and Male (Fig. 5): Head: frons dark brown; eyes chocolate brown covered with

Female (not illustrated): Forewing length 22 mm, n=1. Dorsal surface lighter than male, and ventral dots reflected on dorsal surface. Otherwise differences between male and female are similar to those in the related *L. labda* and *L. lebbaea*.

#### Type

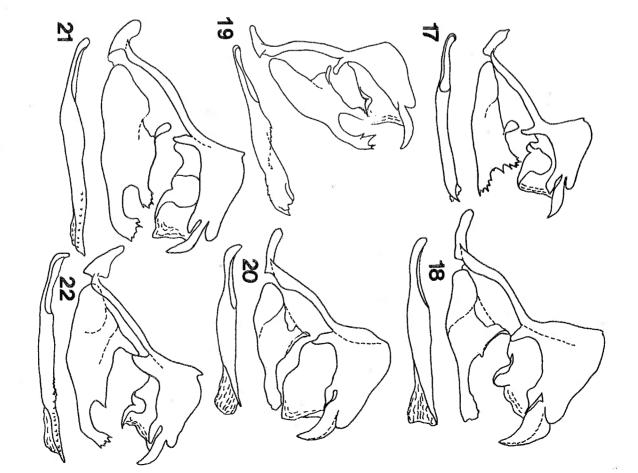
BMNH; 5 males: River Tabaconas, North Peru, 6000 feet, 1912, A. & E. PRATT, male); 4 males: Manchara, North Peru, 7000 feet, IX.1912, A. & E. PRATT, in 2 males: Ecuador, coll. Grose-Smith, in BMNH; 1 male: Ecuador, 1920, coll "Aloag", A. Jasiński leg., in TWP; 2 males: Ecuador, coll. Hewitson, in BMNH 2 males: Tabaconas, Peru, VIII.1996, I. Aldas leg., TWP (1 male), MUSM (1 Brabant, in BMNH; 6 males: Ecuador, no precise data, in TWP. Peru: Cajamarca Pastaza, Ecuador, 3600 feet, M.G. Palmer leg., in BMNH; Vague data: 1 male: San Antonio, 2100 m, IV.1971, Lefebre leg., in BMNH; 1 male: Río Machay, 1550 m, 4-5.VII.93, J. P. W. HALL leg., in KWJH; Pastaza: 1 male: Alpayacu, Río males: Baños, Río Pastaza, 5-7000 feet, M.G. Palmer leg., in BMNH; 1 male: in MNCN (1 male), in PUCE (1 male), in MALUZ (1 male), in TWP (5 males); 13 PB; 8 males: Río Verde Chico, 24.IX.1995, 2100 m, A. Neild & A. Jasniski leg., 06.IV.1995, P. BOYER leg., in PB; 1 male: same locality and collector, V.1996, in 2300 m, 06-07.V.1996, A. Jasiński leg., in TWP; 1 male: Viscaya, 2000 m, Ecuador, 06.IV.1995, local dealer leg., in TWP; 3 males: Biscaya [sic], 2100-San Andrés, Cordillera de Lagunillas, 2000 m, 13.VIII.1998, T. Pyrcz leg. in 15.V-1996, 2500 m, S. Attal & I. Aldas leg., in TWP; 2 males: Biscaya [sic], WILLMOTT leg., in KWJH; 2 males: Viscaya, road to Urba [sic], Baños, Ecuador, TWP; Tunguruhua: 1 male: Ulbilla, Rio Ulba, nr. Baños, 2200 m, 16.XI.96, K. R. Valladolid, National Park Podocarpus, 2700 m, 14.VIII.1998, T. Pyrcz leg., in TWP; 2 males: same locality, V.1998, A. Jasiński leg., in AJ; 2 males: North of 2050 m, 3.XI.96, K. R. WILLMOTT leg., in KWJH; Zamora-Chinchipe: 2 males: Hall leg., in KWJH; Morona-Santiago: 1 male: km. 22 Limón-Gualaceo rd., Catamayo-Porto Velo, Ecuador, 16.II.1993, B. Méry & S. Attal leg., in TWP; PALMER leg., in BMNH. Paratypes (66 males): Ecuador: Loja: 1 male: Route Zamora-Chinchipe: 1 male: north of Valladolid, 2600 m, 18-19.V.94, J. P. W female: Ecuador: Tunguruhua: Baños, Pastaza, east Ecuador, 5-7000 feet, M.G. Ecuador, 22.V.1996, 2200 m, S. Attal & I. Aldas leg., in MZUJ. Allotype Holotype male: Ecuador: Zamora-Chinchipe: Valladolid, Río Chinchipe.

# ETYMOLOGY

This species is dedicated to Nadia Valentina Sánchez, a sister-in-law of the author.

#### REMARKS

Brown (1943) illustrated the male genitalia of *L. nadia* as *L. labda*, pointing out anatomical and colour pattern differences between Ecuadorian (*L. nadia*) and



17-22. Male genitalia: 17 = Lymanopoda nadia n. sp.; 18 - L. caracara n. sp.; 19 - L. nivea bonita n. ssp.;20 - L. ichu n. sp.; 21 - Lymanopoda labineta piniasi n. ssp.; 22 - Lymanopoda excisa browni n. ssp.